



**AssaySense**  
**Human uPA Chromogenic**  
**Activity Kit**

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Thank you for choosing Assaypro.

## Assay Summary

- Step 1.** Add 50  $\mu$ l of Assay Diluent per well.  
Add 30  $\mu$ l of Standard or Sample per well.  
Add 30  $\mu$ l of uPA Substrate per well.
- Step 2.** Read the absorbance at 405 nm for a zero minute background reading.  
Cover and incubate at 37°C.
- Step 3.** Read every 3 minutes for 15 minutes (High Activity).  
Read every 30 minutes for 2 hours (Low Activity).  
Cover and incubate at 37°C after each reading.

## Symbol Key



Consult instructions for use.





# AssaySense Human Urokinase (uPA) Chromogenic Activity Kit (Direct)

Catalog No. CU1001a

*Sample insert for reference use only*

## Introduction

Urokinase-type plasminogen activator (uPA) is a highly restricted serine protease that converts the zymogen plasminogen to active plasmin, a broad-spectrum serine proteinase capable of degrading most of the major protein components of the extracellular matrix. Binding of uPA to its receptor and subsequent uPA mediated pericellular proteolysis are involved in many processes, including cell migration and tissue remodeling in angiogenesis, atherogenesis, tumor cell metastasis, and ovulation (1-2). A high level of uPA is a poor prognostic marker for aggressive breast cancer, aggressive prostate cancer, bladder cancer, and gastric cancer (3-5).

## Principle of the Assay

The AssaySense Human uPA Chromogenic Activity Kit is developed to determine uPA activity in human **plasma, serum, urine, cell culture supernatant, cell lysate, and tissue samples**. The amidolytic activity of uPA is quantitated using a highly specific uPA substrate releasing a yellow para-nitroaniline (pNA) chromophore. The change in absorbance of the pNA at 405 nm is directly proportional to the uPA enzymatic activity.

## Caution and Warning

- This product is for **Research Use Only** and is not intended for use in diagnostic procedures.
- Prepare all reagents, as instructed, prior to running the assay.
- Prepare all samples prior to running the assay. The dilution factors for the samples are suggested in this insert. However, the user should determine the optimal dilution factor.
- The kit should not be used beyond the expiration date.

## Reagents

The activity assay kit contains sufficient reagents to perform 96 tests using the microplate method.

- **Microplate:** A 96-well polystyrene microplate (12 strips of 8 wells).
- **Sealing Tapes:** Each kit contains 3 precut, pressure sensitive sealing tapes that can be cut to fit the format of the individual assay.
- **Human uPA Standard:** Human high molecular weight uPA (10 IU, lyophilized).
- **Assay Diluent (1x):** Buffered protein base (30 ml).
- **uPA Substrate:** Lyophilized, 3 vials.

## Storage Condition

- Upon arrival, immediately store components of the kit at recommended temperatures up to the expiration date.
- Store Standard and uPA Substrate at -20°C.
- Store Microplate and Assay Diluent at 2-8°C.
- Unused microplate wells may be returned to the pouch and resealed.

## Other Supplies Required

- Microplate reader capable of measuring absorbance at 405 nm
- Pipettes (1-20  $\mu$ l, 20-200  $\mu$ l, 200-1000  $\mu$ l, and multiple channel)
- Deionized or distilled reagent grade water
- Incubator (37°C)

## Sample Collection, Preparation, and Storage

- **Plasma:** Collect plasma using one-tenth volume of 0.1 M sodium citrate as an anticoagulant (EDTA or Heparin can also be used as an anticoagulant). Centrifuge samples at 3000 x *g* for 10 minutes and collect plasma. The sample is suggested for use at 1x or within the range of 2x – 5x into Assay Diluent; however, user should determine optimal dilution factor depending on application needs. The undiluted samples can be stored at -20°C or below for up to 3 months. Avoid repeated freeze-thaw cycles.
- **Serum:** Samples should be collected into a serum separator tube. After clot formation, centrifuge samples at 3000 x *g* for 10 minutes and remove serum. The sample is suggested for use at 1x or within the range of 2x – 5x into Assay Diluent; however, user should determine optimal dilution factor depending on application needs. The undiluted samples can be stored at -20°C or below for up to 3 months. Avoid repeated freeze-thaw cycles.
- **Urine:** Collect urine using sample pot. Centrifuge samples at 800 x *g* for 10 minutes. The sample is suggested for use at 1x or within the range of 2x – 5x into Assay Diluent; however, user should determine optimal dilution factor depending on application needs. The undiluted samples

can be stored at -20°C or below for up to 3 months. Avoid repeated freeze-thaw cycles.

- **Cell Culture Supernatant:** Centrifuge cell culture media at 1500 rpm for 10 minutes at 4°C to remove debris and collect supernatant. If necessary, dilute samples into Assay Diluent; user should determine optimal dilution factor depending on application needs. The undiluted samples can be stored at -80°C. Avoid repeated freeze-thaw cycles.
- **Cell Lysate:** The cultured cells are lysed and solubilized with 50 mM Tris-buffered saline (pH 8.0) containing 0.5% Triton X-100 or 0.1% Tween 20 on ice for 30 minutes. Centrifuge samples at 14000 rpm for 10 minutes. Collect fresh cell lysate. If necessary, dilute samples into Assay Diluent; user should determine optimal dilution factor depending on application needs. The undiluted samples can be stored at -80°C. Avoid repeated freeze-thaw cycles.
- **Tissue:** Extract tissue samples with 50 mM Tris-buffered saline (pH 8.0) containing 0.5% Triton X-100 or 0.1% Tween 20 on ice for 30 minutes. Centrifuge samples at 14000 rpm for 20 minutes. Collect supernatant. If necessary, dilute samples into Assay Diluent; user should determine optimal dilution factor depending on application needs. Store remaining extract at -80°C. Avoid repeated freeze-thaw cycles.

*Applicable samples may also include biofluids, cell culture, and tissue homogenates. If necessary, user should determine optimal dilution factor depending on application needs.*

**Refer to Dilution Guidelines for further instruction.**

<b>Guidelines for Dilutions of 100-fold or Greater</b> <i>(for reference only; please follow the insert for specific dilution suggested)</i>	
<b>100x</b>	<b>10000x</b>
A) 4 µl sample : 396 µl buffer (100x) = 100-fold dilution  <i>Assuming the needed volume is less than or equal to 400 µl.</i>	A) 4 µl sample : 396 µl buffer (100x) B) 4 µl of A : 396 µl buffer (100x) = 10000-fold dilution  <i>Assuming the needed volume is less than or equal to 400 µl.</i>
<b>1000x</b>	<b>100000x</b>
A) 4 µl sample : 396 µl buffer (100x) B) 24 µl of A : 216 µl buffer (10x) = 1000-fold dilution  <i>Assuming the needed volume is less than or equal to 240 µl.</i>	A) 4 µl sample : 396 µl buffer (100x) B) 4 µl of A : 396 µl buffer (100x) C) 24 µl of B : 216 µl buffer (10x) = 100000-fold dilution  <i>Assuming the needed volume is less than or equal to 240 µl.</i>

## Reagent Preparation

- Freshly dilute all reagents and bring all reagents to room temperature before use.
- **Human uPA Standard:** Reconstitute the Human uPA Standard (10 IU) with 0.4 ml of Assay Diluent to generate a 25 IU/ml standard stock solution. Allow the vial to sit for 10 minutes with gentle agitation prior to making dilutions.
  - For **high level** uPA activity samples: Prepare duplicate or triplicate standard points by serially diluting from the standard stock solution (25 IU/ml) 2-fold with equal volume of Assay Diluent to produce 12.5, 6.25, 3.125, 1.563, and 0.781 IU/ml solutions. Assay Diluent serves as the zero standard (0 IU/ml).
  - For **low level** uPA activity samples: From the standard stock solution (25 IU/ml), dilute 8-fold with Assay Diluent to produce a 3.125 IU/ml standard working solution. Prepare duplicate or triplicate standard points by serially diluting the standard working solution (3.125 IU/ml) 2-fold with equal volume of Assay Diluent to produce 1.563, 0.781, 0.391, 0.195, and 0.098 IU/ml solutions. Assay Diluent serves as the zero standard (0 IU/ml).

Any remaining stock solution should be stored at -20°C and used within 30 days. Avoid repeated freeze-thaw cycles.

### Standard curve for high level uPA activity samples:

Standard Point	Dilution	[uPA] (IU/ml)
P1	1 part Standard (25 IU/ml)	25
P2	1 part P1 + 1 part Assay Diluent	12.5
P3	1 part P2 + 1 part Assay Diluent	6.25
P4	1 part P3 + 1 part Assay Diluent	3.125
P5	1 part P4 + 1 part Assay Diluent	1.563
P6	1 part P5 + 1 part Assay Diluent	0.781
P7	Assay Diluent	0.0

### Standard curve for low level uPA activity samples:

Standard Point	Dilution	[uPA] (IU/ml)
P1	1 part Standard (25 IU/ml) + 7 parts Assay Diluent	3.125
P2	1 part P1 + 1 part Assay Diluent	1.563
P3	1 part P2 + 1 part Assay Diluent	0.781
P4	1 part P3 + 1 part Assay Diluent	0.391
P5	1 part P4 + 1 part Assay Diluent	0.195
P6	1 part P5 + 1 part Assay Diluent	0.098
P7	Assay Diluent	0.0

- **uPA Substrate:** Add 1.1 ml of reagent grade water to generate a 1x stock solution. Allow the vial to sit for 10 minutes with gentle agitation prior to use. Any remaining stock solution should be stored at -20°C and used within 30 days. Avoid repeated freeze-thaw cycles.

### Assay Procedure

- Prepare all reagents, standard solutions, and samples as instructed. Bring all reagents to room temperature before use. The assay is performed at 37°C in a humid incubator.
- Remove excess microplate strips from the plate frame.
- Add 50 µl of Assay Diluent to each well. Gently tap plate to thoroughly coat the wells. Add 30 µl of Human uPA Standard or sample to each well. Add 30 µl of uPA Substrate to each well. Gently tap plate to ensure thorough mixing. Break any bubbles that may have formed. Read the absorbance at 405 nm for a zero minute background reading. Cover wells with a sealing tape and incubate at 37°C in a humid incubator to avoid evaporation. Incubate microplate at 37°C after each reading.
- For **high level** uPA activity samples, read the absorbance at 405 nm every 3 minutes for 15 minutes.
- For **low level** uPA activity samples, read the absorbance at 405 nm every 30 minutes for 2 hours.

### Data Analysis

- Calculate the mean value of the duplicate or triplicate readings for each standard and sample.
- To generate a standard curve from the optimal reaction time, plot the graph using the standard concentrations on the x-axis and the corresponding mean 405 nm absorbance (OD) or change in absorbance per minute ( $\Delta A/\text{min}$ ) on the y-axis after subtracting the background. The

best fit line can be determined by regression analysis using log-log or four-parameter logistic curve fit.

- Determine the unknown sample concentration from the Standard Curve and multiply the value by the dilution factor.

### Typical Data

- The typical data is provided for reference only. Individual laboratory means may vary from the values listed. Variations between laboratories may be caused by technique differences.

#### Standard data for high level uPA activity samples:

Standard Point	IU/ml	Average OD (15 min)
P1	25	0.673
P2	12.5	0.485
P3	6.25	0.294
P4	3.125	0.178
P5	1.563	0.114
P6	0.781	0.086
P7	0.0	0.053

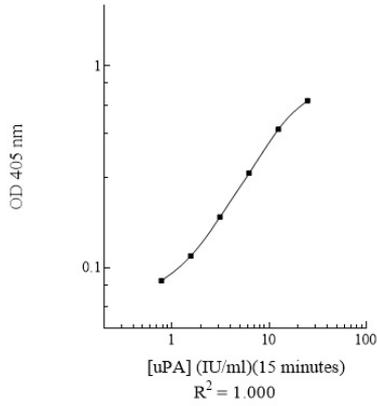
#### Standard data for low level uPA activity samples:

Standard Point	IU/ml	Average OD (2 hours)
P1	3.125	0.652
P2	1.563	0.573
P3	0.781	0.364
P4	0.391	0.217
P5	0.195	0.134
P6	0.098	0.106
P7	0.0	0.051

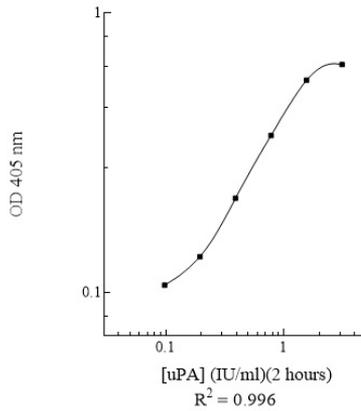
### Standard Curve

- The curve is provided for illustration only. A standard curve should be generated each time the assay is performed.

uPA Chromogenic Activity Standard Curve  
(High uPA Activity Samples)



uPA Chromogenic Activity Standard Curve  
(Low uPA Activity Samples)



## Performance Characteristics

- The minimum detectable dose of human uPA at 2 hours is approximately 0.047 IU/ml.

## Troubleshooting

Issue	Causes	Course of Action
Low Precision	Use of improper components	<ul style="list-style-type: none"> <li>• Check the expiration date listed before use.</li> <li>• Do not interchange components from different lots.</li> </ul>
	Splashing of reagents while loading wells	<ul style="list-style-type: none"> <li>• Pipette properly in a controlled and careful manner.</li> </ul>
	Inconsistent volumes loaded into wells	<ul style="list-style-type: none"> <li>• Pipette properly in a controlled and careful manner.</li> <li>• Check pipette calibration.</li> <li>• Check pipette for proper performance.</li> </ul>
	Insufficient mixing of reagent dilutions	<ul style="list-style-type: none"> <li>• Thoroughly agitate the lyophilized components after reconstitution.</li> <li>• Thoroughly mix dilutions.</li> </ul>
Unexpectedly Low or High Signal Intensity	Microplate was left unattended between steps	<ul style="list-style-type: none"> <li>• Each step of the procedure should be performed uninterrupted.</li> </ul>
	Omission of step	<ul style="list-style-type: none"> <li>• Consult the provided procedure for complete list of steps.</li> </ul>
	Steps performed in incorrect order	<ul style="list-style-type: none"> <li>• Consult the provided procedure for the correct order.</li> </ul>
	Insufficient amount of reagents added to wells	<ul style="list-style-type: none"> <li>• Check pipette calibration.</li> <li>• Check pipette for proper performance.</li> </ul>
	Improper reagent preparation	<ul style="list-style-type: none"> <li>• Consult reagent preparation section for the correct dilutions of all reagents.</li> </ul>
Insufficient or prolonged incubation periods	<ul style="list-style-type: none"> <li>• Consult the provided procedure for correct incubation time.</li> </ul>	
Deficient Standard Curve Fit	Non-optimal sample dilution	<ul style="list-style-type: none"> <li>• User should determine the optimal dilution factor for samples.</li> </ul>
	Contamination of reagents	<ul style="list-style-type: none"> <li>• A new tip must be used for each addition of different samples or reagents during the assay procedure.</li> </ul>
	Contents of wells evaporate	<ul style="list-style-type: none"> <li>• Verify that the sealing film is firmly in place before placing the assay in the incubator or at room temperature.</li> </ul>
	Improper pipetting	<ul style="list-style-type: none"> <li>• Pipette properly in a controlled and careful manner.</li> <li>• Check pipette calibration.</li> <li>• Check pipette for proper performance.</li> </ul>
	Insufficient mixing of reagent dilutions	<ul style="list-style-type: none"> <li>• Thoroughly agitate the lyophilized components after reconstitution.</li> <li>• Thoroughly mix dilutions.</li> </ul>

## References

- (1) Okada S *et al.* (1996) *Arterioscl Thromb Vasc Biol.* 16:1269.
- (2) Besser D *et al.* (1996) *Fibrinolysis.* 10:215.
- (3) Duffy MJ *et al.* (1990) *Cancer Res.* 50:6827.
- (4) Hasui Y *et al.* (1992) *Int J Cancer.* 50:871.
- (5) Nishino N *et al.* (1988) *Thromb Res.* 50:527.

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