



# **Fixation and Permeabilization Buffer Kit**

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# Fixation and Permeabilization Buffer Kit (50 tests)

Catalog No. FIXPERM01

*Sample insert for reference use only*

## Principle of the Assay

The Fixation and Permeabilization Buffer Kit is designed for performing FACS and/or ICC analysis.

## Caution and Warning

- This product is for **Research Use Only** and is not intended for use in diagnostic procedures.
- Prepare all samples prior to starting the procedure.
- The Fixation Buffer contains 4% paraformaldehyde.
- The Permeabilization Buffer contains Triton™ X-100.
- The kit should not be used beyond the expiration date.

## Reagents

- **Fixation Buffer (1x):** A solution to fix cells prior to permeabilization (25 ml, 1 bottle).
- **Permeabilization Buffer (1x):** A surfactant solution to permeabilize cells prior to staining (25 ml, 1 bottle).

## Storage Condition

- Upon arrival, immediately store components of the kit at recommended temperatures up to the expiration date.
- Store Fixation Buffer (1x) and Permeabilization Buffer (1x) at 2-8°C.

## Other Supplies Required

- Flow Cytometer
- Fluorescent Microscope
- Pipettes (1-20 µl, 20-200 µl, and 200-1000 µl)
- 1x PBS, pH 7.4
- Deionized or distilled reagent grade water
- 1x DAPI
- Antifade Mountant

- FACS Buffer (10x): Available for order from Assaypro, catalog no. **FACS**
- Blocking Buffer (10x): Available for order from Assaypro, catalog no. **BB**

## Sample Preparation

- Cells, at about 80% confluence, were collected from 8-10 Petri Dishes/T-75 flasks (2-10 million cells/flask) into conical centrifuge tubes.
- Centrifuge cells at 1300 rpm for 5 minutes at 20°C. Aspirate the supernatant. Wash the pellet once with 10 ml of 1x PBS, pH 7.4 and proceed immediately with the FACS procedure.

## FACS Procedure

- Prepare all reagents and samples as instructed. Keep all reagents at 4°C. The procedure is performed at room temperature (20-25°C). Make sure to resuspend the pellet thoroughly every time after centrifugation.
- Centrifugation takes place for 5 minutes at 1300 rpm and 20°C.  
**Centrifuge and aspirate after each step.**
- Add 2 ml of Fixation Buffer to the prepared sample in the conical centrifuge tubes. Resuspend and incubate for 20 minutes.
- **(Omit this step if you plan to do surface staining)** Wash the pellet two times with 7 ml of FACS Buffer.
- **(Omit this step if you plan to do surface staining)** Add 2 ml of Permeabilization Buffer to the pellet. Resuspend and incubate for 20 minutes.
- Wash the pellet two times with 7 ml of FACS Buffer.
- Add 2 ml of Blocking Buffer to the pellet. Resuspend and incubate for 1 hour.
- Resuspend the pellet in 3 ml of FACS Buffer. Mix the cell suspension thoroughly to make sure there are no visible cell clumps. Aliquot 100 µl of the cell suspension into 5 ml tubes. Add fluorescent conjugated antibody at set concentration (1-2 µg/tube for APC, RPE, and FITC conjugates or 10 µg/tube for PerCP conjugate). Incubate in a dark environment for 45 minutes. Agitate the tube every 5 minutes during incubation to ensure proper mixing or place the tube on a shaker for the entirety of the incubation period.
- Wash the pellet two times with 2 ml of FACS Buffer and centrifuge the tube. Leave 250 µl of supernatant in the tube and aspirate the rest. Add 1 ml of FACS Buffer to resuspend and analyze on a flow cytometer.

## ICC Staining/Corning® Falcon® CultureSlides

- Keep all reagents at 4°C. Make sure cells are at the proper density before proceeding to the next step. **Aspirate after each step.**
- Aspirate media. Add 0.5 ml of 1x PBS to each chamber.
- Add 0.4 ml of Fixation Buffer and incubate for 20 minutes at room temperature. Cover wells during incubation.
- Add 0.5 ml of 1x PBS and incubate for 5 minutes at room temperature. Repeat once.
- Add 0.4 ml of Permeabilization Buffer and incubate for 20 minutes at room temperature.
- Add 0.5 ml of 1x PBS and incubate for 5 minutes at room temperature. Repeat once.
- Add 0.4 ml of Blocking Buffer and incubate for 1 hour at room temperature.
- Add 0.5 ml of 1x PBS and incubate for 5 minutes at room temperature. Repeat once.
- Add fluorescent conjugated antibody to a final volume of 400 µl. Incubate overnight at 4°C.
- Add 0.5 ml of 1x PBS and incubate for 5 minutes at room temperature. Repeat once.
- Stain with 1x DAPI (400 µl/well). Incubate for 4 minutes at room temperature.
- Add 0.5 ml of 1x PBS and incubate for 5 minutes at room temperature. Repeat once.
- Add 25 µl of antifade mountant to each well. Cover with Corning® glass slide cover and incubate overnight at room temperature in a dark environment. **Do NOT aspirate.**
- Analyze using a Fluorescent Microscope.

Version 1.2